

An Inherited Mutation Associated with Functional Deficiency of the α -Subunit of the Guanine Nucleotide-Binding Protein G_s in Pseudo- and Pseudopseudohypoparathyroidism*

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ABSTRACT

Pseudohypoparathyroidism type Ia (PSP) is a disorder characterized by Albright's osteodystrophy, secondary hyperparathyroidism, lowered G_s activity, and resistance of the urinary cAMP excretion to exogenous PTH. The patients had raised basal serum levels of TSH and/or excessive TSH response to TRH. Here we have described a 38-bp deletion at the exon 1/intron 1 boundary of one $G_{s\alpha}$ allele in two mothers with pseudo-PSP and in six offsprings with PSP of a kindred with Albright's osteodystrophy. The deletion eliminates the splice

donor site of exon 1. The pseudo-PSP patients presented decreased G_s activity, but normal urinary cAMP responses to PTH and normal TSH levels and responses to TRH. As monitored during 22 yr, they had normal serum levels of calcium and PTH. The findings demonstrate the same inherited functional defect of $G_{s\alpha}$ in two female patients with pseudo-PSP and in six of their offsprings with PSP. The pathogenesis of clinical hypoparathyroidism remains to be clarified. (*J Clin Endocrinol Metab* 83: 935–938, 1998)

PSEUDOHYPOPARATHYROIDISM Ia (PSP) is a disorder with clinical hypoparathyroidism and the phenotype of Albright's osteodystrophy, but raised plasma levels of bioactive and immunoreactive PTH (1; see Ref. 2 for additional references). Patients with PSP frequently present raised serum levels of TSH and excessive TSH response to TRH with sometimes overt clinical hypothyroidism (3 and references cited therein). The diagnosis of PSP-Ia was established on the basis of resistance of the urinary cAMP excretion to exogenous PTH, and decreased guanine nucleotide-binding protein G_s activity (4–10). G_s activity was decreased in the PSP patients reported here, but it was similarly decreased in their mothers with pseudo-PSP (8). The latter responded normally to exogenous PTH with raised urinary cAMP excretion; secondary hyperparathyroidism and excessive TSH response to TRH were not observed. A functional deficiency of G_s has been reported in PSP and pseudo-PSP (8–10).

A genetic deficiency of $G_{s\alpha}$ has been revealed in familial PSP and pseudo-PSP (11–15; for additional mutations, see Ref. 16). As shown here, the patients with PSP-Ia as well as those with pseudo-PSP carry a novel deletion in one $G_{s\alpha}$ allele resulting in reduced $G_{s\alpha}$ activity. In healthy siblings with no signs of Albright's osteodystrophy, both $G_{s\alpha}$ alleles were normal. It would seem, therefore, that the described genetic defect, resulting in reduced $G_{s\alpha}$ activity, is not an

obvious cause of the resistance to exogenous PTH in the kindred reported here.

Subjects and Methods

Figure 1 shows the pedigree of a family with PSP, pseudo-PSP, and Albright's osteodystrophy. Laboratory data, including serum calcium, serum PTH, urinary cAMP responses to parathyroid extract (Eli Lilly Co., Indianapolis, IN), serum TSH, TSH response to TRH, and G_s protein activity, for patients 7, 9, 14–17, 20, and 22 have been reported previously (3, 5, 8) (Table 1).

Briefly, serum levels of calcium were measured by ethylene glycol-bis (β -aminoethyl ether)- N,N' -tetraacetate titration, using calcein as an indicator, and by flame spectrophotometry (6, 9). Immunoreactive serum PTH was estimated with antibodies to bovine PTH recognizing predominantly intact human PTH-(1–84) in the serum of patients with PSP on gel permeation chromatography. [125 I]Bovine PTH-(1–84) was used as radioligand, and human PTH-(1–84) as standard (8, 17). The urinary cAMP response to parathyroid extract was evaluated according to a slightly modified protocol of Chase *et al.* (4, 5). Immunoreactive TSH and the TSH response to TRH (Hoffmann-La Roche, Basel, Switzerland) were estimated as previously reported (3). G_s protein activity was assessed in erythrocyte ghosts as previously described (8).

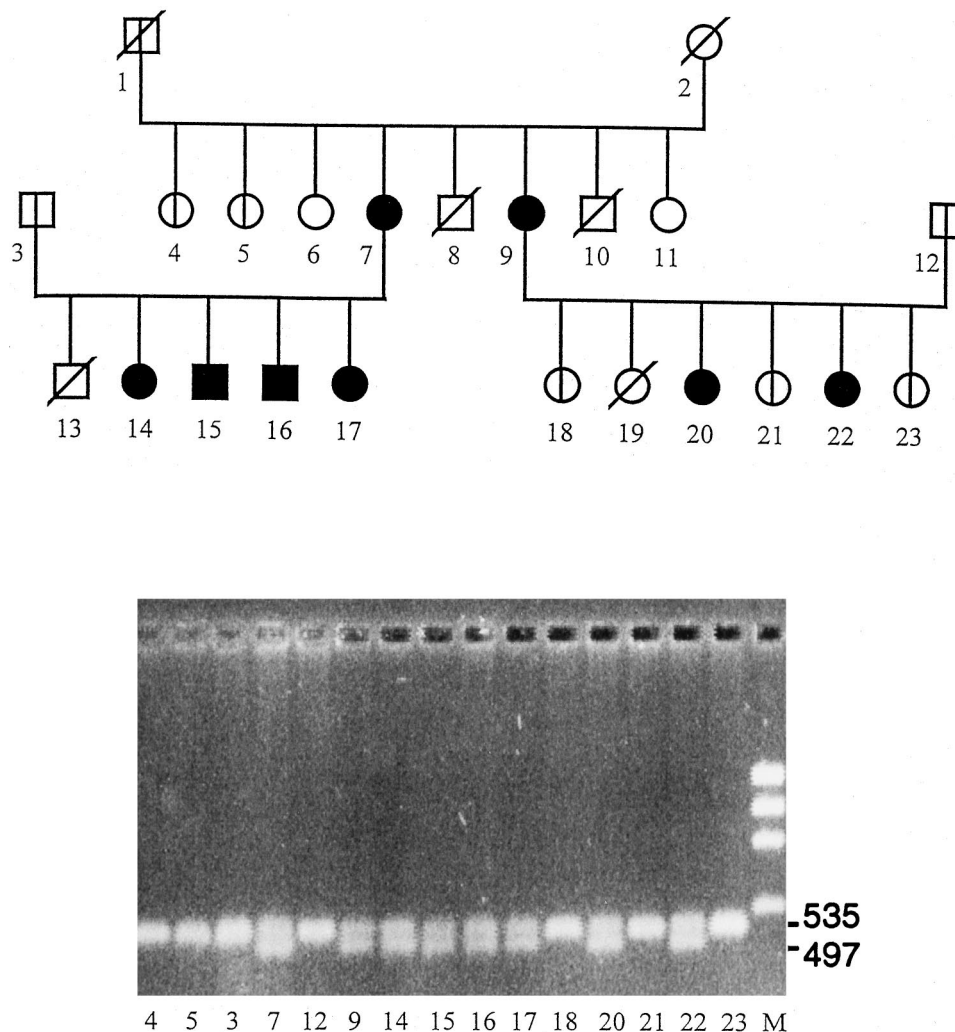
Genomic DNA isolated from peripheral leukocytes of individual subjects was analyzed for $G_{s\alpha}$ gene insertions or deletions by PCR amplification of gene subdomains. The primer pairs indicated in Fig. 2 were used to localize and analyze in detail the deletion described here in exon 1 and flanking regions of the $G_{s\alpha}$ gene. PCR products were separated by electrophoresis in 2% agarose (Seakem HGT, Flowgen Instruments, Sittingbourne, UK) and visualized by

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FIG. 1. Pedigree of a family with (■ and ●) and without (□ and ○) Albright's osteodystrophy. □ and ○, Not tested. Diagonal lines denote deceased subjects (top). Results of agarose gel electrophoresis analysis of DNA products obtained by PCR amplification of DNA isolated from blood samples of selected family members (bottom). The numbers below individual lanes of the agarose gel correspond to the numbers of individual family members, as indicated in the pedigree. M, DNA size markers (ΦX/HaeIII).



ethidium bromide staining. Nucleotide sequence analysis of PCR-amplified $G_s\alpha$ gene fragments was performed by cycle sequencing. The primers used were the same as those used for amplification of gene fragments.

Results

Figure 1 shows the pedigree of the affected family with Albright's osteodystrophy. At first presentation, patients 14–17, 20, and 22 suffered from PSP, with hypocalcemia in four and normocalcemia in two siblings (Table 1). Serum PTH levels were raised, and the patients were classified as PSP-I on the basis of absent or low urinary cAMP responses to exogenous PTH. Five of the six siblings presented raised levels of TSH, and all had excessive TSH response to TRH. G_s activity was decreased in all of them, but G_s was also decreased in patients 7 and 9 with pseudo-PSP exhibiting normal urinary cAMP excretion in response to the administration of parathyroid extract and normal serum levels of TSH and TSH response to TRH at first presentation. They had normal serum levels of calcium and PTH as observed over 22 yr.

Analysis by PCR amplification of exon 1 and flanking regions of the $G_s\alpha$ gene with primers DV-157 and MET1R

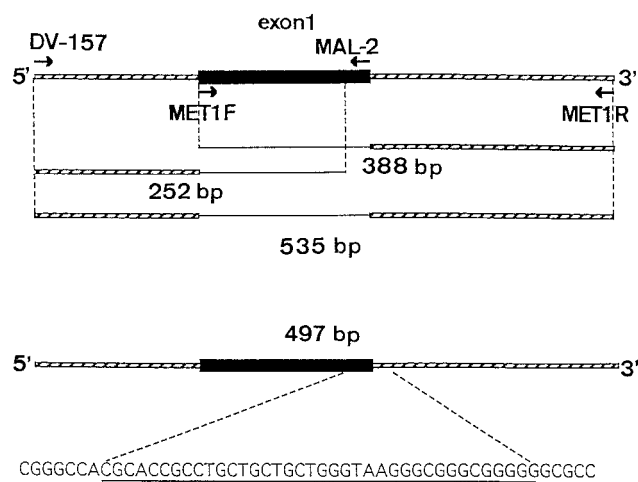
(Fig. 2) revealed a normal-sized 535-bp product in all family members investigated (Fig. 1). In the 6 patients with PSP and the 2 patients with pseudo-PSP, an additional smaller PCR product, indicating a deletion in 1 $G_s\alpha$ allele, was also observed. Subsequent separate PCR and nucleotide sequence analysis of the 5'- and 3'-ends of exon 1 and corresponding flanking regions with primer pairs DV-157/MAL-2 and MET1F/MET1R, respectively, revealed a 38-bp deletion comprising 21 nucleotides of the 3'-end of exon 1 and 17 nucleotides of intron 1 in the mutated allele. This eliminates the donor splice site of exon 1, giving rise to a transcript that includes intron 1. As a result, termination of translation is predicted to occur within intron 1 leading to the incorporation of at least 116 alternative amino acids into a protein product of the mutated $G_s\alpha$ gene.

Discussion

Albright *et al.* (1) made the discovery that PSP is not caused by a lack of PTH, but by an inability to respond to it. This has led to the concept of end-organ resistance to exogenous PTH caused by a defect of the PTH receptor and/or its signaling pathways. Mutations of the PTH/PTHrP receptor have not been detected to date in PSP type Ib (18–20) and have not

Subjects	Age at first presentation (yr)	Serum		cAMP (nmol/min · m ²)		TSH (μU/mL)		G _{sα} (%)
		Calcium (mmol/L)	PTH (ngeq/L)	Basal	Maximal	Basal	Maximal	
1	Deceased (age unknown)	—	—	—	—	—	—	—
2	Deceased (age unknown)	—	—	—	—	—	—	—
3	47	2.27	<100	—	—	—	—	—
4	56	2.24	100	—	—	—	—	—
5	54	2.36	<200	—	—	—	—	—
6	52	—	—	—	—	—	—	—
7	36	2.20	220 ^a	4	90	0.9	13.8	60
8	Deceased (10 days)	—	—	—	—	—	—	—
9	34	2.35	135 ^b	4	226	1.7	13.0	55
10	Deceased (1.17 yr)	—	—	—	—	—	—	—
11	42	—	—	—	—	—	—	—
12	49	2.45	<200	—	—	—	—	—
13	Deceased (age unknown)	—	—	—	—	—	—	—
14	16	2.05	1840	1.1	11.6	13.1	25.0	—
15	6	1.58	341	2.7	6.0	2.0	28.9	50
16	5	1.35	390	0.2	5.1	12.2	18.8	47
17	13	2.00	480	3.1	9.4	9.5	33.2	—
18	21	2.30	281	—	—	—	—	—
19	Deceased (1 yr)	—	—	—	—	—	—	—
20	5	2.43	390	3.8	1.4	7.0	32.4	39
21	26	2.31	327	—	—	—	—	—
22	1	2.45 ^a	270 ^c	4.7	11.0	7.6	39.4	51
23	12	2.36	<200	—	—	—	—	—
Control subjects (range)		2.08–2.43	<300	0.3–5.9	17–268	<1.0–3.3	10.0–18.0	91–113

^c At 13 yr, serum calcium was 2.35 mmol/L, and PTH was 1640 ng/L.



been reported in PSP type Ia. The findings that urinary cAMP excretion in response to exogenous PTH is reduced or absent and of an excessive TSH response to TRH are consistent with inadequate activation of the PTH- and TRH-responsive adenyl cyclase and different isotypes thereof, and activation of phosphodiesterase (4, 16). To this end, G_s activity was shown to be reduced (7–11). Subsequently, several mutations of the G_{α} -encoding gene have been discovered (12–16). The

Circulating bioactive and immunoreactive PTH levels are normal or raised in PSP, but not in pseudo-PSP (2). Plasma from patients with PSP revealed higher PTH inhibitory activity, assessed in a renal cytochemical bioassay, than that in their mothers with pseudo-PSP presented here (22). Intact PTH was separated from a putative inhibitor on gel permeation chromatography of plasma samples. A postulated PTH antagonist whose structure remains to be elucidated may be responsible for the renal resistance to PTH observed in PSP. Yet, some patients with PSP and secondary hyperparathyroidism have osteitis fibrosa (23).

In conclusion, the hypothesis of target organ resistance to PTH being caused by an inactivating mutation of one $G_s\alpha$ allele is questionable in the kindred reported here.

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References

- Albright F, Burnett CH, Smith PH, Parson W. 1942 Pseudohypoparathyroidism—an example of "Seabright-Bantam syndrome." *Endocrinology*. 30:922–932.
- Nagant de Deuxchaisnes C, Fischer JA, Dambacher MA, et al. 1981 Dissociation of parathyroid hormone bioactivity and immunoreactivity in pseudohypoparathyroidism type I. *J Clin Endocrinol Metab*. 53:1105–1109.
- Werder E, Illig R, Bernasconi S, et al. 1975 Excessive thyrotropin response to thyrotropin-releasing hormone in pseudohypoparathyroidism. *Pediatr Res*. 9:12–16.
- Chase LR, Melson GL, Aurbach GD. 1969 Pseudohypoparathyroidism: defective excretion of 3',5'-AMP in response to parathyroid hormone. *J Clin Invest*. 48:1832–1844.
- Werder EA, Fischer JA, Illig R, et al. 1978 Pseudohypoparathyroidism and idiopathic hypoparathyroidism: relationship between serum calcium and parathyroid hormone levels and urinary cyclic adenosine-3',5'-monophosphate response to parathyroid extract. *J Clin Endocrinol Metab*. 46:872–879.
- Farfel Z, Brickman AS, Kaslow HR, Brothers VM, Bourne HR. 1980 Defect of receptor-cyclase coupling protein in pseudohypoparathyroidism. *N Engl J Med*. 393:237–242.
- Levine MA, Downs Jr RW, Singer M, Marx SJ, Aurbach GD, Spiegel AM. 1980 Deficient activity of guanine nucleotide regulatory protein in erythrocytes from patients with pseudohypoparathyroidism. *Biochem Biophys Res Commun*. 94:1319–1324.
- Fischer FA, Bourne HR, Dambacher MA, et al. 1983 Pseudohypoparathyroidism: inheritance and expression of deficient receptor-cyclase coupling protein activity. *Clin Endocrinol (Oxf)*. 19:747–754.
- Levine MA, Jap T-S, Mauseth RS, Downs RW, Spiegel AM. 1986 Activity of the stimulatory guanine nucleotide-binding protein is reduced in erythrocytes from patients with pseudohypoparathyroidism and pseudopseudohypoparathyroidism: biochemical, endocrine, and genetic analysis of Albright's hereditary osteodystrophy in six kindreds. *J Clin Endocrinol Metab*. 62:497–502.
- Saito T, Akita Y, Fujita H, et al. 1986 Stimulatory guanine nucleotide binding protein activity in the erythrocyte membrane of patients with pseudohypoparathyroidism type I and related disorders. *Acta Endocrinol (Copenh)*. 111:507–515.
- Levine MA, Ahn TG, Klupt SF, et al. 1988 Genetic deficiency of the α subunit of the guanine nucleotide-binding protein G_s as the molecular basis for Albright hereditary osteodystrophy. *Proc Natl Acad Sci USA*. 85:617–621.
- Patten JL, Johns DR, Valle D, et al. 1990 Mutation in the gene encoding the stimulatory G protein of adenylate cyclase in Albright's hereditary osteodystrophy. *N Engl J Med*. 322:1412–1419.
- Weinstein LS, Gejman PV, Friedman E, et al. 1990 Mutations of the G_s α -subunit gene in Albright hereditary osteodystrophy detected by denaturing gradient gel electrophoresis. *Proc Natl Acad Sci USA*. 87:8287–8290.
- Farfel Z, Iiri T, Shapira H, Roitman A, Mouallem M, Bourne HR. 1996 Pseudohypoparathyroidism, a novel mutation in the $\beta\gamma$ -contact region of $G_s\alpha$ impairs receptor stimulation. *J Biol Chem*. 271:19653–19655.
- Iiri T, Farfel Z, Bourne HR. 1997 Conditional activation defect of a human $G_{s\alpha}$ mutant. *Proc Natl Acad Sci USA*. 94:5656–5661.
- Levine MA. 1996 Pseudohypoparathyroidism. In: Bilezikian JP, Raisz LG, Rodan GA, eds. *Principles of bone biology*. San Diego: Academic Press; 853–876.
- Loveridge N, Tschopp E, Born W, Devogelaer J-P, Nagant de Deuxchaisnes C, Fischer JA. 1986 Separation of inhibitory activity from biologically active parathyroid hormone in patients with pseudohypoparathyroidism type I. *Biochim Biophys Acta*. 889:117–122.
- Suarez F, Lebrun JJ, Lecossier D, Escoubet B, Coureau C, Silve C. 1995 Expression and modulation of the parathyroid hormone (PTH)/PTH-related peptide receptor messenger ribonucleic acid in skin fibroblasts from patients with type Ib pseudohypoparathyroidism. *J Clin Endocrinol Metab*. 80:965–970.
- Schipani E, Weinstein LS, Bergwitz C, et al. 1995 Pseudohypoparathyroidism type Ib is not caused by mutations in the coding exons of the human parathyroid hormone (PTH)/PTH-related peptide receptor gene. *J Clin Endocrinol Metab*. 80:1611–1621.
- Fukumoto S, Suzawa M, Takeuchi Y, et al. 1996 Absence of mutations in parathyroid hormone (PTH)/PTH-related protein receptor complementary deoxyribonucleic acid in patients with pseudohypoparathyroidism type Ib. *J Clin Endocrinol Metab*. 81:2554–2558.
- Schwindinger WF, Reese KJ, Lawler AM, Gearhart JD, Levine MA. 1997 Targeted disruption of *G-nas* in embryonic stem cells. *Endocrinology*. 138:4058–4063.
- Loveridge N, Fischer JA, Nagant de Deuxchaisnes C, et al. 1982 Inhibition of cytochemical bioactivity of parathyroid hormone by plasma in pseudohypoparathyroidism type I. *J Clin Endocrinol Metab*. 54:1274–1275.
- Kolb FO, Steinbach HL. 1962 Pseudohypoparathyroidism with secondary hyperparathyroidism and osteitis fibrosa. *J Clin Endocrinol Metab*. 22:59–64.